

## Chemoenzymatic Synthesis of (2*S*,3*R*)-3-Hydroxyproline from Cyclopentadiene

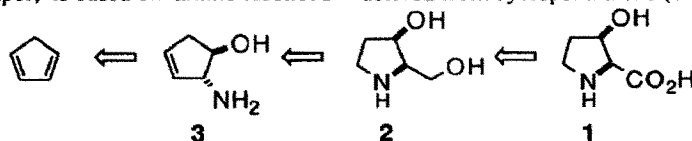
Hari Sundram, Adam Golebiowski, and Carl R. Johnson\*

Department of Chemistry, Wayne State University, Detroit, Michigan 48202-3489

**Abstract:** Amido-alcohol **4**, derived from cyclopentadiene and resolved using *Candida antarctica* lipase B-mediated transacetylation, was transformed to (2*R*, 3*R*)-3-hydroxy-2-hydroxymethylpyrrolidine (**2**) and (2*S*, 3*R*)-3-hydroxyproline (**1**).

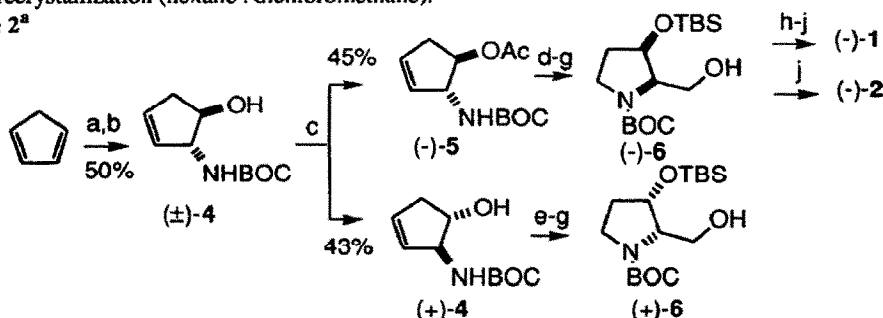
(2*S*, 3*R*)-3-Hydroxyproline (**1**) is a known, although rare,  $\alpha$ -amino acid found in teleomycin antibiotics.<sup>1,2</sup> It can be easily transformed into the Geissman-Waiss lactone<sup>3</sup> and subsequently into a variety of pyrrolizidine alkaloids,<sup>3, 4</sup> including retronecine, platynecine, and croalbinecine. (2*S*, 3*R*)-3-Hydroxyproline (**1**) and (2*R*, 3*R*)-3-hydroxy-2-hydroxymethylpyrrolidine (**2**) are present as structural units in pyrrolizidine alkaloids (e.g., australine<sup>5</sup> and alexine<sup>6</sup>), indolizidine alkaloids (e.g., slaframine,<sup>7</sup> castanospermine<sup>8</sup>) and unusual amino acids (e.g., detoxinine<sup>9</sup>). Several syntheses of *cis*-3-hydroxyproline have been reported.<sup>4, 10</sup> A new approach, presented in this paper, is based on amino alcohol **3**<sup>11</sup> derived from cyclopentadiene (Scheme 1).

Scheme 1



Enzymatic resolution of racemic alcohol **4** was achieved by *Candida antarctica* lipase B (Novo Nordisk SP 435)-mediated transacetylation in isopropenyl acetate to afford acetate (-)-**5** in 45 % yield (>98% ee) and alcohol (+)-**4** in 43% yield (93% ee) (Scheme 2).<sup>12</sup> The enantiopurity of alcohol (+)-**4** was increased to >98% by a single recrystallization (hexane : dichloromethane).

Scheme 2<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) AcOOH then NH<sub>3</sub>/MeOH (ref.11); (b) BOC<sub>2</sub>O, EtOAc; (c) *Candida antarctica* lipase B (Novo Nordisk SP 435), isopropenyl acetate, RT, 4 h; (d) KOH, MeOH (97%); (e) TBSCl, imidazole, DMF (98%); (f) O<sub>3</sub>, MeOH - CH<sub>2</sub>Cl<sub>2</sub> (1 : 1), then DMS; (g) NaBH<sub>3</sub>CN, AcOH, MeOH (f + g: 71%); (h) NaOCl, TEMPO, KBr, NaHCO<sub>3</sub>, Et<sub>2</sub>O-H<sub>2</sub>O; (i) NaIO<sub>4</sub>, RuCl<sub>3</sub> (cat.), Me<sub>2</sub>CO (h + i: 56%); (j) HCl, MeOH-H<sub>2</sub>O (1: 91%; 2: 88%).

Processing of (-)-**5** by protective group exchange (O-Ac → O-TBS),<sup>13</sup> followed by ozonolysis and sodium cyanoborohydride reduction gave known prolinol (-)-**6**.<sup>10c,14</sup> Acidic hydrolysis afforded pyrrolidine **2**, while stepwise oxidation<sup>15,16</sup> (TEMPO, NaOCl then RuCl<sub>3</sub>, NaIO<sub>4</sub>) led, after deprotection, to 3-hydroxyproline (**1**).<sup>14</sup> The enantiomeric 3-hydroxyprolinol derivative (+)-**6**<sup>14</sup> was obtained from alcohol (+)-**4**.<sup>14</sup>

This chemoenzymatic method allows access to both enantiomers of 3-hydroxyproline (**1**) and the related 3-hydroxy-2-hydroxymethylpyrrolidines from cyclopentadiene. The lipase-mediated acetylation/resolution in organic media provides convenient access to enantiopure intermediates (+)- and (-)-**4** which should prove useful in the synthesis of a variety of enantiopure targets including pyrrolizidine and indolizidine alkaloids.

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12. The enzymatic acetylation of alcohol ( $\pm$ )-**4** was conducted as follows: alcohol ( $\pm$ )-**4** (500 mg, 2.6 mmol) was dissolved in isopropenyl acetate (3 mL) and to this solution SP-435 (immobilized recombinant *Candida antarctica* lipase B) (50 mg, 10 % by wt.) was added. The reaction progress was monitored by GC. At ca. 50% conversion (approx. 4.5 h) the reaction mixture was filtered, concentrated and chromatographed on silica gel (hexane : ethyl acetate; 5:1) to give acetate (-)-**5** (225 mg, 45% yield, 98% ee. by  $^{19}\text{F}$  NMR of the Mosher ester) and alcohol (+)-**4** (260 mg, 43% yield, 93% ee. by  $^{19}\text{F}$  NMR of the Mosher ester).  
For examples of other applications of the SP-435 biocatalyst from our laboratory see: Johnson, C. R.; Braun, M. P. *J. Am. Chem. Soc.* **1993**, *115*, 11103; Johnson, C. R.; Bis, S. *J. Tetrahedron Lett.* **1992**, *33*, 7287; Johnson, C. R.; Sakaguchi, H. *Synlett* **1992**, 813.
13. This exchange resulted in improved yields in the subsequent TEMPO oxidation.
14. Physical Data: **1** as HCl salt: mp 225 °C d.;  $[\alpha]_{\text{D}}^{23}$  -95.5 (c 0.5, H<sub>2</sub>O) [lit. 10b,  $[\alpha]_{\text{D}}^{20}$  -101 (c 1.0, H<sub>2</sub>O)];  $^1\text{H}$  NMR as reported.<sup>10f</sup>  
**2** as HCl salt: mp 220 °C d.;  $[\alpha]_{\text{D}}^{23}$  - 12.7 (c 0.2, MeOH);  $^1\text{H}$  &  $^{13}\text{C}$  NMR as reported.<sup>17</sup>  
( $\pm$ )-**4**: mp 76.0-77.0 °C;  $^1\text{H}$  NMR (CDCl<sub>3</sub>)  $\delta$  5.75 (dd, J = 5.7, 2.1 Hz, 1H); 5.45 (d, J = 3.9 Hz, 1H); 5.00 (bs, 1H); 4.32 (bs, 1H); 4.15 (m, 2H); 2.65 (ddd, J = 17.1, 7.2, 1.8 Hz, 1H); 2.17 (dd, J = 17.1, 2.4 Hz, 1H); 1.36 (s, 9H);  $^{13}\text{C}$  NMR (CDCl<sub>3</sub>)  $\delta$  156.6, 132.7, 128.2, 79.8, 79.7, 65.5, 39.4, 28.2.  
(-)-**5**: mp 71.0-72.0 °C;  $[\alpha]_{\text{D}}^{23}$  -107.4 (c 0.7, CHCl<sub>3</sub>);  $^1\text{H}$  NMR (CDCl<sub>3</sub>)  $\delta$  5.85 (m, 1H); 5.78 (m, 1H); 5.07 (m, 1H); 4.70-4.55 (m, 2H); 2.86 (dd, J = 17.7, 6.9 Hz, 1H); 2.27 (bd, J = 17.7 Hz, 1); 2.06 (s, 3H); 1.44 (s, 9H);  $^{13}\text{C}$  NMR (CDCl<sub>3</sub>)  $\delta$  155.1, 131.8, 129.7, 79.6, 62.0, 38.0, 28.3, 21.1.  
(+)-**4**: mp 72.0-73.0 °C;  $[\alpha]_{\text{D}}^{23}$  +19.1 (c 1.0, CHCl<sub>3</sub>).  
(-)-**6**: oil;  $[\alpha]_{\text{D}}^{23}$  -35.4 (c 3.1, CHCl<sub>3</sub>) [lit. 10c,  $[\alpha]_{\text{D}}^{25}$  -34.4 (c 1.9, CHCl<sub>3</sub>)].  
(+)-**6**: oil;  $[\alpha]_{\text{D}}^{23}$  +33.5 (c 5.0, CHCl<sub>3</sub>)
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